

# The Structure and Population Genetics of the Breakpoints Associated With the Cosmopolitan Chromosomal Inversion *In(3R)Payne* in *Drosophila melanogaster*

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## ABSTRACT

We report here the breakpoint structure and sequences of the *Drosophila melanogaster* cosmopolitan chromosomal inversion *In(3R)P*. Combining *in situ* hybridization to polytene chromosomes and long-range PCR, we have identified and sequenced the distal and proximal breakpoints. The breakpoints are not simple cut-and-paste structures; gene fragments and small duplications of DNA are associated with both breaks. The distal breakpoint breaks the *tolkin* (*tok*) gene and the proximal breakpoint breaks CG31279 and the *tolloid* (*tld*) gene. Functional copies of all three genes are found at the opposite breakpoints. We sequenced a representative sample of standard (*St*) and *In(3R)P* karyotypes for a 2-kb portion of the *tok* gene, as well as the same 2 kb from the pseudogene *tok* fragment found at the distal breakpoint of *In(3R)P* chromosomes. The *tok* gene in *St* arrangements possesses levels of polymorphism typical of *D. melanogaster* genes. The functional *tok* gene associated with *In(3R)P* shows little polymorphism. Numerous single-base changes, as well as deletions and duplications, are associated with the truncated copy of *tok*. The overall pattern of polymorphism is consistent with a recent origin of *In(3R)P*, on the order of  $N_e$  generations. The identification of these breakpoint sequences permits a simple PCR-based screen for *In(3R)P*.

THROUGH the classic work of Dobzhansky on *Drosophila pseudoobscura*, chromosome inversions claim an important place in the early study of genetic variation in natural populations (POWELL 1997). Because inversions reduce recombination in heterokaryotypes, they are a genomic feature with potential to come under natural selection and play a role in evolution. A common belief is that inversion polymorphisms are maintained by balancing selection (DOBZHANSKY 1970). If so, individual inversions might be ancient, that is, old relative to a hypothetical neutral arrangement (ANDOLFATTO *et al.* 2001). This age hypothesis can be examined by comparing patterns and levels of sequence variation between and within inverted and standard (*St*) arrangements. Inverted regions are subject to reduced recombination. On a timescale relevant to the question of balancing selection, however, recombination is often not sufficiently suppressed by the inversions to prevent exchanges (see ASHBURNER 1989; HASSON and EANES 1996). Any assessment of inversion age using a sequence-derived estimate of gene genealogy will, therefore, be confounded

by crossing over and gene conversion operating within the inverted arrangement (HASSON and EANES 1996). However, topological constraint on homolog pairing increases with increasing proximity to the inversion breakpoints (NOVITSKI and BRAVER 1954). This increase in constraint results in decreasing crossing over with increasing proximity to the breakpoints. For this reason, recovering nucleotide sequences as close to the breakpoints as possible offers the most informative way to study the population genetics of inversion polymorphisms. Finally, the structural features at the breakpoints may also offer insight into the molecular nature and mutational origin of the inversion (*e.g.*, transposable elements), the potential for genetic damage by disrupting gene function, and the genealogical uniqueness of the arrangement.

*Drosophila melanogaster* possesses well-studied cosmopolitan inversion polymorphisms on all four autosomal arms, as well as a common X-linked inversion in east African populations (KRIMBAS and POWELL 1992). In 1994, using a mixed strategy of chromosome microdissection, nonspecific PCR, and reciprocal hybridization to a *St* arrangement genomic library, WESLEY and EANES (1994) reported the molecular characterization of the breakpoints of the *In(3L)Payne* inversion in *D. melanogaster*. The *In(3L)P* breakpoints are basic cut-and-paste structures. No novel features, such as transposable elements or other arrangements, were associated with the breakpoints. A population study of sequence variation indicated that the arrangement, while not ancient, also appeared to be not recent; it emerged from the base of the inferred genealogy of *St* arrangements and pos-

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essed several fixed differences from the *St* arrangement. In 1999, the *In(2L)t* inversion was extensively studied by ANDOLFATTO and colleagues (ANDOLFATTO *et al.* 1999; ANDOLFATTO and KREITMAN 2000). They recovered a sequence spanning the proximal breakpoint and reported on a large population-based sample of sequences >5 kb in length. They predicted that the inversion was not old and estimated the minimum age to be  $0.3N$  generations or  $\sim 100,000$  years. There was an unusual haplotype structure associated with the *St* chromosomes in this region, as well as an apparent excess of molecular polymorphism in the breakpoint region, all suggesting balancing selection.

In this report, we describe the breakpoints of a third *D. melanogaster* cosmopolitan inversion *In(3R)Payne*, using the known cytological position and *D. melanogaster* genome sequence as a starting reference point for a long-range PCR walk. We find breakpoints with complex arrangements, including duplication and fragmentation of the *tolkin* (*tok*) gene. Sequence data for the *St* arrangement copies of *tok*, and the inverted copy of *tok* and its partial pseudogene, predict a relatively recent origin of this cosmopolitan inversion.

## MATERIALS AND METHODS

**Long-range PCR and *in situ* hybridization:** Earlier cytological studies predicted that the breakpoints of *In(3R)P* are at 89C2–3 (proximal breakpoint) and 96A18–19 (distal breakpoint, KRIMBAS and POWELL 1992). To identify the precise molecular breakpoints we used long-range PCR and *in situ* hybridization of amplified fragments to *In(3R)P*-bearing chromosomes and focused on the 89C2–3 breakpoint. Primers were synthesized using the sequence of the *D. melanogaster* genome, release 3.2.2 (ADAMS *et al.* 2000; <http://www.fruitfly.org>), and starting from a site in the region of the predicted (cytological) breakpoint. The strategy was to move successively closer to the breakpoint until the long-range, and eventually short-range PCRs failed. We utilized two lab-created lines homozygous for *In(3R)P* and a single-standard line (Bloomington *Drosophila* Stock Center line 2057). Under this strategy the breakpoint sequence would eventually be recovered via inverse PCR using standard methods (OCHMAN *et al.* 1988). Briefly, a region of known sequence was used to design reverse orientation PCR primers and to identify suitable restriction endonuclease sites. Genomic DNA from a *In(3R)P* line was digested with various endonucleases, self-ligated to form loops, and used as template for PCR reactions. This method is efficient at recovering regions of unknown sequence that are contiguous with regions of known sequence. Long-range PCR was carried out using the Expand Long Template PCR system (Roche Applied Science, Mannheim, Germany). Primers were designed to the specifications of the polymerase enzyme system. *In situ* hybridization of polytene chromosomes was performed following a modified version of the Berkley *Drosophila* Genome Project protocol (<http://www.fruitfly.org/aout/methods/cytogenetics.html>). PCR fragments ( $\sim 10$  kb) were labeled with digoxigenin (Roche Applied Science) and *in situ* hybridization carried out on both the standard line and a line homozygous for *In(3R)Payne*.

**Populations:** Iso-third chromosome lines were collected in 1997 and have been characterized in other studies (DUVER-

NELL AND EANES 2000; VERRELLI AND EANES 2001; SEZGIN *et al.* 2004). In the population study reported here, lines with *In(3R)P* were identified by a PCR-based screen using nested primers. A set of three PCR primers was designed from the above work to screen for the inversion: 12253917+ (ACT AGC GTT GAG AAT GCA AAG TCC AAC), 12254223– (AAA TGC TGC ACG TAA TTG TAA GTT ATG AGC), and 20560888– (TTT GTT TGT GTC TGT GTG AGC TGC). The numbering again refers to the annotated *D. melanogaster* genome sequence, release 3.2.2 (ADAMS *et al.* 2000). Given the duplication associated with the inversion (see RESULTS), primer pair 12253917+ / 12254223– will amplify a 306-bp fragment in both *St* and inverted chromosomes, while primer pair 12254223– / 20560888– will amplify a 663-bp fragment only from inverted chromosomes. The accuracy of this method was tested by PCR screening 10 *St* and 10 inverted isochromosomal lines previously karyotyped using the salivary gland preparations. All chromosome karyotypes were correctly scored using the PCR screen.

We amplified and sequenced an  $\sim 2$ -kb fragment, starting at the distal breakpoint, from both *St* and *In(3R)P* chromosomes. All *In(3R)P* lines available from the earlier studies were sequenced (SEZGIN *et al.* 2004). Sequenced *St* lines were selected to match the number and location of the *In(3R)P* lines used. Samples spanned collection sites from southern Florida to Maryland. The amplified  $\sim 2$ -kb fragment covers a segment of the *tolkin* gene (*tok*) in the standard chromosomes and the beginning of the *tok* pseudogene in *In(3R)P* chromosomes. We also sequenced the same  $\sim 2$ -kb fragment from the functional copy of the *tok* gene in *In(3R)P* chromosomes. PCR primers designated 20560313+ (TTG GCC TAA TCG AAT TGC TAT G) and 20562431– (CGC CAC CGC AAA CAA CTA TGG) were used to amplify a 2.1-kb fragment from both the *St* orientation chromosomes and the functional copy on the *In(3R)P* chromosomes. PCR primers 12254223– and 20562431– amplified a similarly sized fragment of the *tok* gene broken by the *In(3R)P* inversion. Six lines of *Drosophila simulans*, initially collected on Long Island, New York, full-sibling inbred for 20 generations, and the not-yet-annotated *D. simulans* genome sequence (<http://www.genome.wustl.edu/projects/simulans/index.php>) were used for sequence comparison to root the phylogenetic analysis and to determine the likely ancestral state of polymorphic sites.

**Sequencing and data analysis:** PCR amplification products were cleaned using a QIAGEN PCR purification kit and sequenced directly using both the initial PCR primers and the internal primers. All sequencing was done by the DBS sequencing facility, University of California at Davis, using Big Dye Terminator version 3.1 (ABI) and sequenced on a 3730 DNA analyzer (ABI). Base calls from the chromatograms were checked, and the 2-kb fragments were assembled, using Sequencher (Gene Codes, Ann Arbor, MI). The *St tok* and *In(3R)P* inverted *tok* and pseudogene *tok* fragment were aligned using Clustal X (THOMPSON *et al.* 1997), with minor adjustments done by eye, resulting in a final aligned sequence of 2053 bases. All sequences have been deposited in GenBank (accession nos. AY881252–AY881292).

The effective number of synonymous and nonsynonymous sites was estimated using DnaSP 4.0 (ROZAS AND ROZAS 1997). This program was also used to calculate  $\theta$  and  $\pi$  and carry out the TAJIMA (1989) and FU AND LI (1993) tests. A phylogeny was constructed using the neighbor-joining method (SAITOU AND NEI 1987) as implemented in MEGA version 2.1 (KUMAR *et al.* 2001) and the Kimura two-parameter model (KIMURA 1980) for distance estimation. Gaps in the aligned sequence were deleted in a pairwise manner and 1000 bootstrap replicates

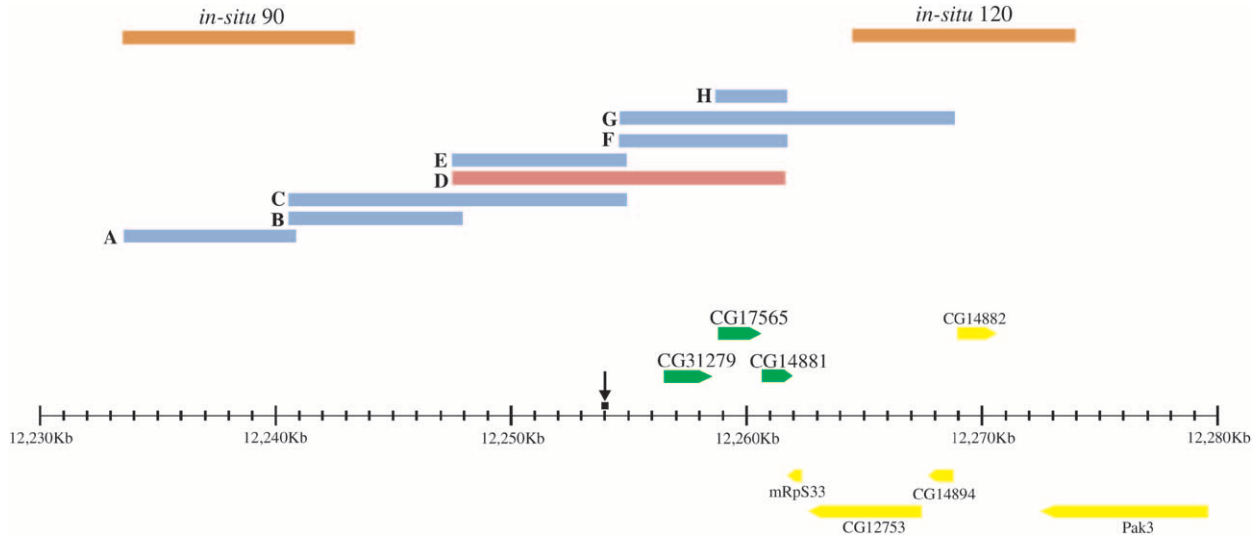


FIGURE 1.—Proximal region of chromosomal inversion *In(3R)P* (89C2–7). The ruler indicates the position in arm 3R according to the *D. melanogaster* genome. The numbers in parentheses indicate the numbering scheme used for the primers. Coding genes immediately flanking the inversion breakpoints (see Figure 2) are illustrated in green; other coding genes covered by the *in situ* and long-range PCR analysis are shown in yellow. Blue bars indicate the locations of PCR amplifications that worked in both inverted and standard lines. The red bar indicates the location of the PCR amplification that worked only in standard lines. Orange bars indicate the locations of the *in situ* probes. The black arrow indicates the 289-bp region implicated in our long-range PCR screen to contain the proximal breakpoint. Primers used for A were (12233497+, 1240696–), B (12240514+, 12247904–), C (12240514+, 12254800–), D (12247448+, 12261570–), E (12247448+, 12254800–), F (12254500+, 12261570–), G (12254500+, 12268649–), and H (12258824+, 12261719–). The 90 and 120 *in situ* probes were created using primers 12233497+/12243306 and 12264288+/12273660, respectively.

(FELSENSTEIN 1985) performed to evaluate support for each node in the tree.

## RESULTS

### Defining the breakpoints: Localization of breakpoints:

Figure 1 shows the region of the proximal breakpoint, its predicted genes, and the genomic fragments (between 3 and 14 kb) amplified by long-range PCR in the initial search for the *In(3R)P* inversion breakpoints. The numbering refers to the annotated *D. melanogaster* genome sequence, release 3.2.2 (ADAMS *et al.* 2000; <http://www.fruitfly.org>). *In situ* hybridization of the 90 and 120 fragments (Figure 1) to polytene chromosomes of *In(3R)P* confirmed that the breakpoint fell between 12,243 and 12,264 kb in the *D. melanogaster* genome sequence, but all long-range PCR amplifications across this 20-kb region worked on both *St* and *In(3R)P* samples. One explanation for this paradox was a possible duplication in the *In(3R)P* chromosome of the region containing the positive primers for fragments G and F and/or the negative primers for E and C. An ~14-kb fragment (marked D) failed to amplify in inverted arrangements, but amplified in *St* chromosomes. Our approach was to carry out subsequent smaller PCR reactions, while maintaining one of the primers of D constant. Using the same negative primer as in D (12261570–, CAT TAT ACC AGC CAC CAC CGC GTT GAG CAG G), we designed positive prim-

ers that would amplify smaller pieces. None of these smaller PCR reactions should work in an inverted sample unless the positive primer was within the duplicated region (all PCR reaction will amplify from a standard line). We observed that PCR amplification failed when using primer 12253902+ (ACT AGC GTT GAG AAT GCA AAG TCC AAC), but was successful when utilizing 12254192+ (GTT GCT CAT AAC TTA CAA TTA CGT GCA GC). This suggested that the end of the duplicated region (the breakpoint) was between these two positive primers (Figure 1; black bar and arrow).

To localize the distal breakpoint, reverse orientation primers, 12254224– (AAA TGC TGC ACG TAA TTG TAA GTT ATG AGC) and 12254500+ (TGA TGC AGT CCG ACG ACA ACA ACA CAA GCA GC) were designed next to this 289-bp region to allow inverse PCR amplification of the flanking DNA. These primers amplified an ~500-bp fragment in *TaqI*-digested and self-ligated DNA from a *In(3R)P* line. Sequencing this fragment we found sequence from coordinate 12,254 kb contiguous with exon 4 of the *tokin* gene (*tok*; NGUYEN *et al.* 1994; FINELLI *et al.* 1995). The *tok* gene lies near the predicted (cytological) distal breakpoint, while 12,254 kb is near the predicted proximal breakpoint. A second inverse PCR, using *MspI* digested *In(3R)P* DNA, yielded an ~400-bp fragment. Sequencing this fragment we again found bases from 12,254 kb contiguous with the *tok* fourth exon. The junction of sequences from the region of the

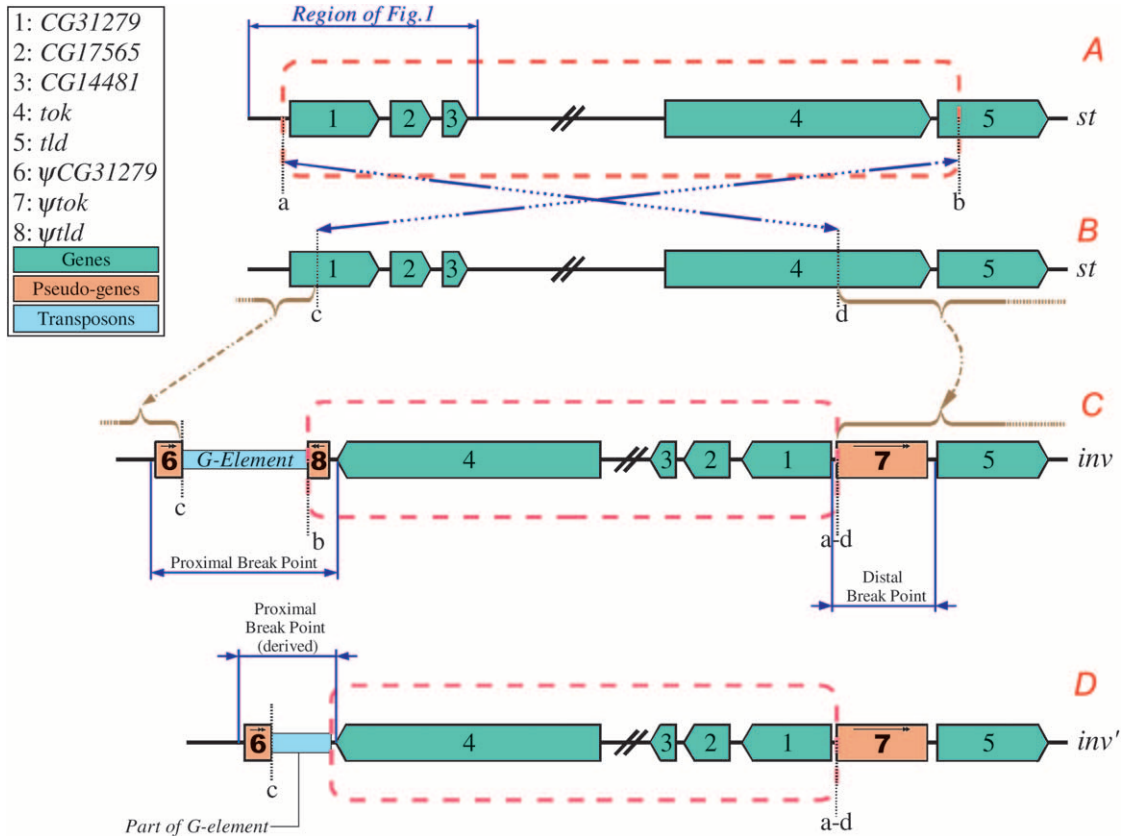


FIGURE 2.—Structure of breakpoints for *Standard* and *In(3R)P* chromosomes. Both the distal and proximal *In(3R)P* breakpoints split, and presumably inactivate, known coding genes. Each “split” gene is asymmetrically duplicated at the opposite breakpoint. The boxed text indicates the numbering and color-coding scheme. (A and B) Standard orientation chromosomes showing the inversion breakpoints and putative pattern of the inversion event. (C and D) Inverted chromosomes. The chromosome in D contains a small deletion at the proximal breakpoint and is presumably derived from C. See RESULTS and Figure 3 for a detailed description of the breakpoints. This figure is not to scale.

cytological predicted proximal and distal breakpoints appears to have identified the distal breakpoint of *In(3R)P*. This breakpoint appears to break the *tok* gene. To further clarify and confirm the distal breakpoint, we attempted to PCR amplify sequence between a primer in the *CG31279* gene, the nearest coding gene to the genomic sequence recovered in the inverse PCR (*i.e.*, inside the inversion), 12255485– (CAG CAG CAG CTA CTT GGC TTT TAT TTA TT), and a primer located distal to the predicted distal breakpoint, 500-bp past the 3'-end of the *tok* gene 20565305– (CCT TAG GCA TCT TAG CAT AAG TCA ATG GGT GGC). As predicted, this amplification failed in *St* lines, but produced an ~6-kb fragment in *In(3R)P* lines. This fragment was completely sequenced from three lines. The fragment contained bases 12,255,485–12,254,041 of the *D. melanogaster* genome sequence contiguous with 4471 bp of the *tok* gene and 434 bases past the 3'-end of the *tok* gene. The *tok* sequence included 269 bases of exon 4, and all of the downstream region (3'-end) of the *tok* gene; exons 1–3 and the first 18 bases of exon 4 are not present. This *tok* fragment is most likely a pseudogene ( $\psi$ tok, block 7 in Figures 2 and 3). The *tok* end of this fragment

matches the *St* chromosome sequence and it appears that this 7-kb amplification crosses the distal breakpoint.

*Characterization and sequencing of the breakpoints:* We next attempted to amplify and sequence the proximal breakpoint. To begin, we conducted inverse PCR on *In(3R)P* and *St* samples using primers designed to match the portion of the *tok* gene missing at the distal breakpoint of *In(3R)P* chromosomes. If the proximal breakpoint was simply a cut-and-paste match to the distal breakpoint, these primers would amplify a fragment of unknown size across the proximal breakpoint of the *In(3R)P* samples. The primers were expected to also amplify a 750-bp fragment from the functional copy of *tok* in *St* chromosome. This amplification, however, produced a 750-bp fragment from *TaqI*-digested DNA from both *In(3R)P* and *St* samples, suggesting that a second copy of the *tok* gene existed in the *In(3R)P* chromosome.

Since our initial long-range PCR results suggested the possibility of a duplication in the region of the inversion breakpoints, a second copy of the *tok* gene was not unexpected. Furthermore, loss-of-function mutations of the *tok* gene are homozygous lethal (FINELLI *et al.* 1995), yet *In(3R)P* homozygotes are viable. Our finding that

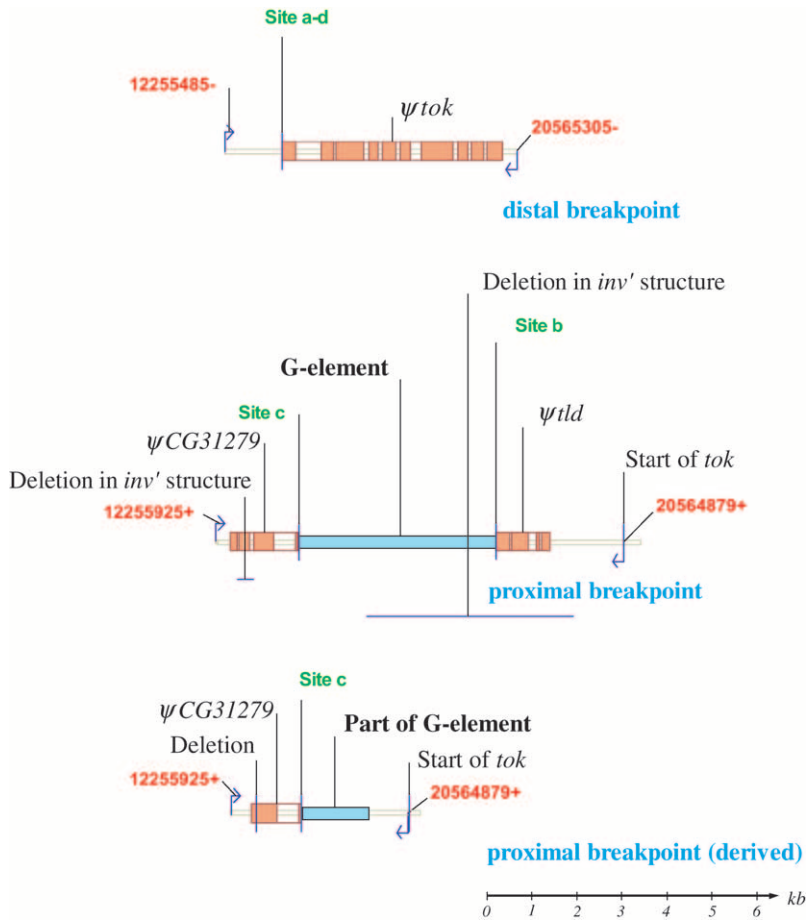


FIGURE 3.—Detail of the distal, proximal, and derived proximal breakpoints from the *In(3R)P* chromosomes. Numbering and color coding are as in Figure 2. Primers and primer sites from the long-range PCR used to characterize each breakpoint are indicated. See RESULTS for details. A scale bar is shown in the bottom right-hand corner.

the distal breakpoint of *In(3R)P* fragmented the *tok* gene suggested that either the *tok* gene was duplicated elsewhere or some compensatory change in another functionally connected gene was involved. If, however, a duplicate copy of *tok* did exist it would have to be closely linked to either breakpoint to keep the inversion from being homozygous lethal. To test whether a second copy of the *tok* gene was associated with the proximal breakpoint, we attempted to amplify from a primer at the 3'-end of the *tok* sequence, 20564879+ (TCA GTG GTT CCA CCA TAG CC), to a primer in the *CG31279* gene, 12255925+ (TGA GTT TCG GCA TAA ATT ACG A). The primers were oriented such that amplification would be possible only if a duplicated copy of *tok* was in an inverted orientation (our expectation if it were a product of the inversion that led to *In(3R)P*) and located near a copy of *CG31279* (in the *St* orientation). The primers were designed such that amplification was not possible from the copy of *CG31279* and  $\psi tok$  fragment found at the distal breakpoint.

This PCR amplification yielded an  $\sim 7$ -kb fragment, both ends of which were sequenced. The fragment crossed from the 3'-end of the *tok* gene into the 5'-end of *CG31279*, presumably crossing the proximal breakpoint (Figure 2, blocks 6 and 8; Figure 3). Sequencing 2.5 kb of the *tok* end of the fragment revealed 1339 bp of the

intergenic region between *tok* and *tld* and exons 1–4 of the *tld* gene, suggesting that a functional copy of the *tok* gene is associated with the proximal breakpoint. Continuing along the fragment, the *tld* sequence is contiguous with sequence that appears to be from a *G*-element-like transposable element (DI NOCERA *et al.* 1986; KAMINKER *et al.* 2002). Sequencing 2 kb of the “CG31279 end” of this 7-kb fragment, we found all of the exons 1–3, and 18 bp of exon 4, of *CG31279*. The *CG31279* sequence is then contiguous with sequence from a *G*-element-like transposable element. The truncated *tld* and *CG31279* are most likely pseudogenes (designated  $\psi tld$  and  $\psi CG31279$ ). The *CG31279* end of the fragment matches the *St* chromosome sequence and we are confident that this 7-kb amplification crosses the proximal breakpoint. The presence of *G*-element-like sequence at the ends of both  $\psi tld$  and  $\psi CG31279$  suggests that such an element separates these two fragments in the proximal breakpoint of *In(3R)P*. The size and restriction digest of a fragment amplified from this putative element also match those expected of a *G* element (see below). The duplicated primer sites that confounded the initial long-range PCR walk are found in the regions of the  $\psi CG31279$ ,  $\psi tld$ , and *tok* sequences at the proximal breakpoint. Further characterization of both breakpoints is diagrammed in Figure 3.

The mutational event that resulted in the *In(3R)P* arrangement left a complicated genomic structure consisting of duplicated coding sequences and pseudogene fragments at each breakpoint. We hypothesize that *In(3R)P* might have arisen by inversion of a region of a *St* chromosome and replacement of a smaller chromosomal region of its sister chromosome. This would have resulted in duplication of the regions flanking both breakpoints and prevented the fragmentation of genes at the breakpoints from being lethal mutations. This model is diagrammed in Figure 2; coding genes are shown in green, and pseudogenes are shown in orange. The red broken-line box outlines the region of the *St* chromosome that was inverted, the proximal and distal ends of the inverted region are denoted a and b, respectively (Figure 2A). The insertion sites are indicated in Figure 2B and denoted c and d, and the arrows between Figures 2A and 2B indicate the inversion. The *In(3R)P* structure is shown in Figure 2C, and the likely origin of the pseudogene fragments created by inversion and replacement are indicated by the arrows between Figures 2B and 2C. In the *In(3R)P* chromosome, one copy of CG31279 is broken at site c leaving a 1391-bp fragment of exons 1–4 ( $\psi$ CG31279, block 6 in Figure 2; Figure 3). The inversion also breaks the *tld* gene at site b, leaving 1114 bp of exons 1–4 ( $\psi$ tld, block 8 in Figures 2 and 3).

When we sequenced the proximal breakpoint in different *In(3R)P* lines we observed two slightly different breakpoint structures. In one structure (Figure 2C; Figure 3), a *G*-element-like sequence is found between  $\psi$ CG31279 and  $\psi$ tld (between blocks 6 and 8). In the other structure (Figure 2D; Figure 3, *inv'*), which is likely a derivative of the first structure, the  $\psi$ tld gene was not present and only 1352 bp of the 5'-end of the *G* element remained. It appears that this second structure resulted from an imprecise excision of the *G* element from the breakpoint resulting in the concurrent deletion of the  $\psi$ tld fragment. The inverted (functional) copy of the *tok* gene is still intact in this derived structure. Eight of the 13 *In(3R)P* proximal breakpoints sequenced had the longer sequence structure (Figure 3, *inv'*), the remaining five had the shorter, apparently derived, structure (Figure 3, *inv'*). All five lines with the *inv'* proximal breakpoint structure also had a 354-bp deletion that covers all of the second exon of their  $\psi$ CG31279 sequence (Figure 3). The sequences of the distal, proximal, and derived-proximal breakpoints have been submitted to GenBank (accession nos. AY886890–AY886892).

We did not sequence the entire  $\sim$ 3.5 kb of the *G*-element region of the *In(3R)P* structure, just  $\sim$ 300 bp of the ends. These ends exactly match both ends of a transposable element sequence on the *X* chromosome (bases 44576–47033 of GenBank submission AC011704) and are 73% identical to the canonical *G*-element sequence (DI NOCERA *et al.* 1986; KAMINKER *et al.* 2002). Additionally, the length of the long-range PCR amplification

fragment, from  $\psi$ CG31279 to  $\psi$ tld (between blocks 6 and 8, Figure 2), implied that the element size of the insertion is approximately the same as the *X* chromosome *G*-element-like sequence. Restriction endonuclease digestions with *EcoRI* and *BspHI* of a 4-kb fragment amplified from this putative element matches the expected digestion pattern of this *X* chromosome *G*-element-like sequence. There is no transposable element insertion at the distal breakpoint of *In(3R)P* in any of the lines in our study.

**Sequence variation:** We amplified and sequenced 2050 bases of the single *tok* in *St* lines and both the functional *tok* and  $\psi$ tok from *In(3R)P* lines. Two earlier studies characterized the genomic structure of the *tolkin* gene (NGUYEN *et al.* 1994; FINELLI *et al.* 1995); our fragment begins at the nineteenth base of exon 4 (this is the first base after the distal breakpoint and defines the 5'-end of  $\psi$ tok). The first 519 amino acids of the TOK protein form an N-terminal proregion that is apparently proteolytically cleaved to produce a functional TOK protein (NGUYEN *et al.* 1994; FINELLI *et al.* 1995). In our fragment, bases 1–270 and bases 792–834 code for the last 104 amino acids of this proregion. We have separated these bases from the rest of the coding regions in our analysis of sequence polymorphism because such regions are known to evolve under very different constraints than “typical” coding regions (GARCIA-MAROTO *et al.* 1991). Our analysis also covers all of exons 5–7, introns 4–6, and 84 bases of intron 7. We chose to study population level variation in the *tok* gene because it is better annotated than CG31279 and the pseudogene fragment was present in a greater number of *In(3R)P* lines than the *tld* fragment.

We sequenced representative regions of the *St tok* gene and the functional *tok* and  $\psi$ tok copies found in *In(3R)P* using lines from our eastern seaboard populations (DUVERNELL and EANES 2000; VERRELLI and EANES 2001; SEZGIN *et al.* 2004). The region included 1790 bp common to 13 copies of the *St* arrangement *tok* gene, 13 copies of the functional *In(3R)P tok* gene, and 9  $\psi$ tok copies from *In(3R)P*. Four copies of *In(3R)P* would not amplify for the  $\psi$ tok region and are presumed to bear large insertions. The variable sites are listed in Figure 4, and Figure 5 depicts a neighbor-joining tree (SAITOU and NEI 1987) to show general relationships among the sequences. Table 1 summarizes the statistics associated with these regions. The *tok* gene from *St* arrangements shows normal levels of synonymous polymorphism ( $S = 15$ ;  $\theta = 0.022$ ) for *D. melanogaster* genes (MORIYAMA and POWELL 1996). The inverted (functional) *tok* gene in the *In(3R)P* arrangement shows very little synonymous polymorphism ( $S = 1$ ;  $\theta = 0.001$ ). Between the *St* and *In(3R)P* there are five fixations in the functional *tok* gene, but these are not the result of acquired mutations since origination. All five changes are present in both functional *tok* and  $\psi$ tok sequences in *In(3R)P* and there-

	11122	2333333334	4555555567	7778999999	9000111112	1111111111	1111111111	1111111111	1111
	125603345	7001223461	5133469970	0774022235	7557335772	4570245679	2377890246	6139	
	7119875759	9452552066	4078177811	2574034563	1193250142	2367646118	2123129160	2357	
Genome	GTGCAACGCT	T-TTCCCTCT	AAAT-G--TT	A-CACTC-AT	TGTTATCTAC	GGTCGTGTCT	AT--GCCCCC	AAGG	
MD45	.CA.....	..C1.T-A	.....	.....	.....	.....	...A...	.G..	
JFL13	.C.GG....	..C1.T-A	.....	.....	.....	..C....TG	.CAT.7.T.	.G..	
VA4	.CA.G.GA..	..C1.T-A	.....	.....	.....	..CT....TG	.CAT.7.TGT	.G..	
VA5	.C.GG.GA..	..C1.T-..	..T-.TC.A	..T.T-..	.....	..C....TG	.C--.A...	.G..	
MD55	..G.T....	G-A.1T....	..T-.TC.A	..T.T-..	.....	..C....TG	.C--.A...	.G..	
NC12	.C.GG.GA..	.CA.1T....	<b>2</b> .T-.TC..	..T....-C	CC.....	.....	...A...	.G..	
NC19	..G.T....	G-A.1T....	..T-.TC..	T.T....-C	CC.....	..A.....	...A...	.G..	
MFL47	..G.T....	GCA.1T....	..T-.TC..	..T....-C	CC.....	..C....TG	.CAT.7.TGT	G..A	
MFL63	.CA.....	..1.....	..T-.TC..	..T.T-..	CC..CCACGT	..C....TG	.C--.A...	..G..	
MFL67	.CA.....	..C1.....	..T-.TC..	..T.T-..	CC..CCACGT	..C....TG	.C--.A...	.G..	
NC8	.CA.....	..1.....	..T-.TC..	..T.T-..	CC..CCACGT	..C....TG	.C--.A...	.G..	
VA10	.CA.....	..1.....	..T-.TC..	..T.T-..	CC..CCACGT	..C....TG	.C--.A...	.G..	
MFL1	.CA.....	..1.....	..T-.TC..	..T.T-..	CC..CCACGT	..C....TG	.C--.A...	.G..	
MFL61	.CA....TC	G-.1-....	...-TC..	..T....-C	CC.....T	.C..C..G	.C--.A...	..G..	
JFL5	.CA....TC	G-.1-....	...-TC..	..T....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
JFL84	.CA....TC	G-.1-....	..C-TCC..	..T....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
MFL42	.CA....TC	G-.1-....	..C-TCC..	..T....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
HFL116	.CA....TC	G-.1-....	...-TC..	..T....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
VA20	.CA....TC	G-.1-....	...-TC..	..T....-C	CC.A....T	.C..C..G	.CAT.7...T	.G..	
MFL44	.CA....TC	G-.1-....	...-TCC..	..T....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
NC43	.CA....TC	G-.1-....	...-TC..	..T....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
MFL69	.CA....TC	G-.1-....	...-TC..	..T....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
HFL8	.CA....TC	G-.1-....	...-TC..	..T....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
MD49	.CA....TC	G-.1-....	...-TC..	..T....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
MFL62	.CA....TC	G-.1-....	...-TC..	..T....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
NC5	.CA....TC	G-.1-....	..C-TCC..	..T....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
ψ MFL69	.CA....TC	G-.1-....	...-TC..	..T....-C	CA.A....T	.C..C..G	.CATT7....	.G..	
ψ JFL84	.CA....TC	G-.1-....	...-TC..	..T....-C	CA.A....T	.C..C..G	.CATT7....	.G..	
ψ MFL61	.CA....TC	G-.1-.G.	..T-.TC..	..T....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
ψ MD49	.CA....TC	G-.1-....	...-TC..	..T....-C	CC6-----T	.AC..C..G	GCATT7....	.GA.	
ψ NC43	.CA....TC	G-.1-....	<b>3</b> .TC..	..T....-C	CC.A....T	.AC..C..G	GCATT7....	.G..	
ψ HFL8	.CA....TC	G-.1-....	...-ATC..	<b>TT</b> ....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
ψ NC5	.CA....TC	G-.1-....	...-ATC..	<b>TT</b> ....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
ψ VA20	ACA....TC	G-.1-....	...-TC..	..T....-C	CC.A....T	.C..C..G	.CATT7....	.G..	
ψ MFL62	.CA....TC	G-.1-....	...-TC..	..T..CG45-	-C.A....T	A.C..CA..G	.CATT7....	.G..	
D. Sim	.CA....C	G-.1-....	...-TC..	..T....-C	CC..C..CG.	.C..C..TG	.C--.7....	.G.T	

FIGURE 4.—Variable sites for a 2053-bp fragment of the *tokin* gene from the *St* chromosomes (first 13 sequences) and the corresponding regions from both the functional (second 13 sequences) and inactivated ( $\psi$ ) *tok* genes in *In(3R)P* chromosomes (see Figures 2 and 3). Base 1 of this fragment is the first base distal to the distal breakpoint. Coding sites are highlighted: polymorphic sites in the proregion of the protein are highlighted gray; those in the rest of the coding sequence are highlighted blue. Nonsynonymous changes are shown in boldface type. The underlined change is to a stop codon. The boldface numbers refer to insertion/deletion events: 1, a variable length poly(C) string; 2, a 12-bp deletion; 3, a 17-bp insertion; 4, an 11-bp insertion; 5, a 46-bp deletion; 6, a 142-bp deletion; 7, an 8-bp deletion. Red stars indicate the sites unique to the *In(3R)P* sequences. The reference sequence (“Genome”) is from the annotated *D. melanogaster* genome, release 3.2.2 (ADAMS *et al.* 2000; <http://www.fruitfly.org>).

fore reflect differences in the founding *tok* allele at the time of duplication and origin of the inversion. Furthermore, the functional *tok* gene copy from line MFL61 is exceptional. It possesses several polymorphisms that are common to the *St* arrangement and appears to be a recombination (gene conversion) product. This apparent gene conversion event further inflates the estimate of *S* in the *In(3R)P* copies of *tok*. The truncated  $\psi$ *tok* copy associated with the inversion breakpoint shows many features expected of a nonfunctional copy (Figure 4). Of the 10 copies, 5 possess insertions and deletions that would disrupt function. All of the TAJIMA (1989) and

FU and LI (1993) tests conducted on the *tok* sequences from constructed sets of *St* and *In(3R)P* chromosomes were statistically nonsignificant.

### DISCUSSION

The breakpoints of *In(3R)P* are not the simple two cut-and-paste structures such as observed in the *In(3L)P* inversion (WESLEY and EANES 1994). The duplication of several kilobases adjacent to the breakpoints was unexpected and confounded our initial attempt to PCR walk across the distal breakpoint. We would not have rec-

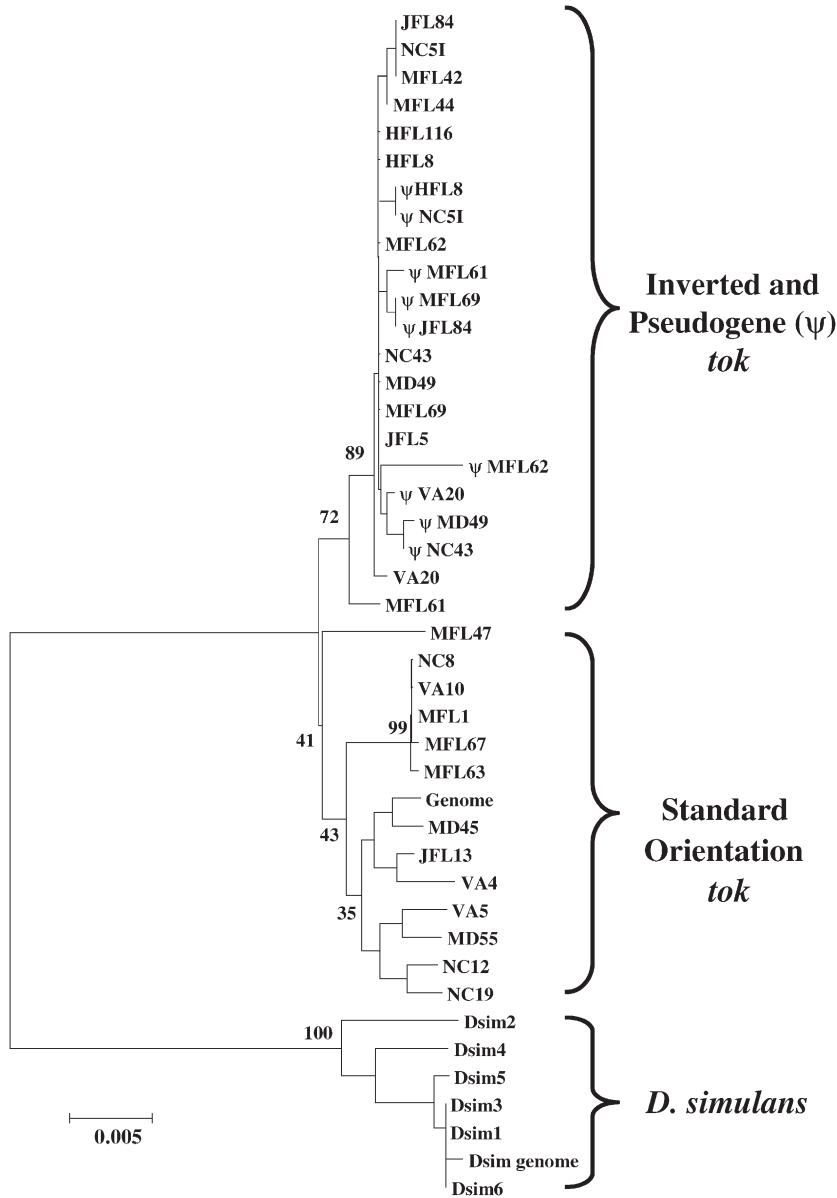


FIGURE 5.—Neighbor-joining tree of *D. melanogaster tok* sequences rooted with the sequences from *D. simulans*. Numbers at key nodes represent the proportion of 1000 bootstrap replicates supporting that node.

ognized that we had passed over the breakpoint had it not been for the parallel confirmation of the breakpoint region using *in situ* chromosome hybridization with flanking probes. The placement of our long-range primers coincidentally landed in the duplicated regions. The precise molecular mechanism for the inversion event is not apparent, although unequal exchange between two sister chromosomes seems to be a reasonable explanation. It is noteworthy that a *G* element is found at the proximal breakpoint in all the inverted copies. Whether the *G* element existed before the inversion event, or has inserted after the fact, cannot be determined from our data. The associated duplications add another twist to the story of inversion breakpoints and the origin of inversions in *Drosophila* and *Anopheles*, where previously both simple cut-and-paste and transposable elements have been implicated in breakpoints (CIRERA *et*

*al.* 1995; MATHIOPOULOS *et al.* 1998; CACERES *et al.* 2001; CASALS *et al.* 2003).

One inverted *tok* sequence MFL61 contributes entirely to the sharing of any polymorphisms between *St* and *In(3R)P* arrangements. This shared tract probably represents a gene conversion event. While reciprocal exchange recombination will be severely reduced in such close proximity to the breakpoint, gene conversion is not (ROZAS and AGUADÉ 1994). We also observed short-gene conversion tracts in our study of the *Pgm* gene, which is located just inside the proximal breakpoint of *In(3L)P* (VERRELLI and EANES 2000).

The relative low level of polymorphism of functional *tok* in *In(3R)P* could also be the signature of a young age for the inversion, although the possibility that *In(3R)P* has historically been low in frequency would also explain this observation. The observation of extensive polymor-



TABLE 1

Comparison of levels of polymorphism ( $\pi$  and  $\theta$ ) within and between *St* and *In(3R)P* chromosomes for the three regions of the *tokin* locus studied

	Bases <sup>a</sup> :	312	725	955
	Proregion <sup>b</sup>		Introns	Exons
		Standard		
<i>S</i>		7	17	16
Non		8	NA	1
Syn		2	NA	15
No. of sites		66.15	725	215.74
$\pi$		0.012	0.008	0.031
$\theta$		0.007	0.007	0.022
Indels		0	7	0
		Inverted		
<i>S</i>		0	6	1
Non		0	NA	0
Syn		0	NA	1
No. of sites		66.33	725	215.83
$\pi$		0	0.002	0.001
$\theta$		0	0.003	0.001
Indels		0	3	0
		Pseudogene		
<i>S</i>		1	4	7
Non		1	NA	5
Syn		0	NA	2
No. of sites		66.33	725	181.04
$\pi$		0	0.002	0.004
$\theta$		0	0.002	0.004
Indels		0	3	2

Non, nonsynonymous changes; syn, synonymous changes.

<sup>a</sup> The number of sites used in each calculation, excluding positions with gaps. For exons, the number of sites,  $\pi$ , and  $\theta$  were all calculated for synonymous sites only.

<sup>b</sup> The first 519 amino acids of the TOK protein, 104 of which are included in our analysis, are proteolytically cleaved from the functional protein.

phism in the  $\psi tok$  copies, which are linked to the functional *tok* sequences, argues against recent strong bottlenecks or adaptive sweeps. The frequency of *In(3R)P* varies widely; it is absent from most northern populations, yet can be the majority arrangement in some tropical and subtropical populations (KRIMBAS and POWELL 1992; SEZGIN *et al.* 2004). We can estimate the historical population frequency and subsequent minimum age of the inversion (in units of  $N_e$  generations) using the ratio of polymorphisms as in ANDOLFATTO *et al.* (1999). This estimation assumes that upon origination the *In(3R)P* inversion rapidly moved to its current frequency and has subsequently reached mutation-drift neutral equilibrium. Ignoring the clear case of MFL61 as a gene conversion event, we observe only four polymorphic mutations in the functional *tok* sequences from our sample of 12 *In(3R)P* chromosomes and 40 in our sample of 13 *St* chromosomes. From these numbers and associated sample sizes we estimate the historical

frequency of *In(3R)P* to be 0.091, and the *minimum* age to be  $0.33N_e$  generations. This is the time back to the most recent common ancestor (MRCA) of the *In(3R)P* sequences. Failure to reach mutation-drift equilibrium will cause us to underestimate the historical frequency; however, this estimate of historical frequency is consistent with the global frequency of the inversion, so the low polymorphism *per se* is not necessarily an indication of recent origin.

The failure to find fixed *de novo* differences in the *tok* sequence variation among the *St* and inverted sequences strongly suggests that *In(3R)P* is not an ancient inversion polymorphism (predating the MRCA ancestor of *St* chromosomes). The number of fixed differences between arrangements can also be used to provide an independent estimate of age. There are five fixed differences, but the sharing of all these differences between *tok* and  $\psi tok$  shows that none have arisen since the inversion event. From a coalescence perspective, fixations within the inversion lineage must have occurred prior to the MRCA of *In(3R)P* and the MRCA with the *St* arrangement. There are no true fixed differences, so this defines the lower time of *In(3R)P* origination to *In(3R)P* MRCA as simply “recent.” The upper time interval depends on the maximum time interval over which there is a probability of no fixations having occurred by chance under Poisson sampling. We can put some confidence on this upper interval of time.

The average level of divergence in the *D. melanogaster* lineage (since the MRCA with *D. simulans*) is 0.0852 changes per silent site (DUNN *et al.* 2001), and TAMURA *et al.* (2004) estimate the time of divergence between *D. melanogaster* and *D. simulans* as 5.4 million years. For a region with 231 silent sites the expected divergence would be  $\sim(0.085 \times 231) = 19.68$  changes or  $19.68/2.7 = 7.288$  changes/region/million years. Assuming a Poisson distribution of changes per unit time, a time interval with expected mean of 3.0 mutations has a 5% chance of realizing a sample of no fixed mutations under Poisson sampling. Therefore, an upper 95% age confidence interval for the age of the inversion would be  $3.00/7.288 = 411,000$  years. This translates into  $4.11 \times 10^6$  generations, using a reasonable estimate for *D. melanogaster* of  $\sim 10$  generations/year. With the widely used estimate of  $N_e \approx 10^6$  for *D. melanogaster* (KREITMAN 1983) this suggests a total upper age limit of  $4.11 N_e$  and a lower age limit of  $0.33N_e$  generations (see above). Therefore, it is possible that the *In(3R)P* inversion dates to the MRCA of the *St* arrangements. It is also worth noting that of the five “fixed” mutations captured by the inversion, two are the ancestral state (shared with *D. simulans*), suggesting that the inversion is old enough for those states to now not be represented in the contemporary *St* arrangements. Nevertheless, our observations for *In(3R)P* are consistent with the assertion by ANDOLFATTO *et al.* (2001) that most inversion polymorphisms

have ages on the order of  $N_e$  generations and are not ancient.

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